Extraction of Acacia senegal MYB transcription factor mRNA by using homology searching approach: An important tree species of arid environment

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Abstract

Acacia senegal one of the most dominant biomass producer of the sandy areas of the desert, belonging to (Fabaceae family and genus Acacia) commonly known as Kummat. These plants has well known to contain high levels and different types of secondary metabolites (gum Arabic), which are often responsible for their therapeutic and pharmacological activities. In-group of transcription factors of plants, MYB (myeloblastosis) gene family most diversified in various aspects. The study was to finding MYB genes sequences by using homological approaches. Applied standardized method for isolation and purification of genomic DNA from Acacia senegal leaves followed by an amplification of MYB genes. Primers has designed using online Genefisher2 and Primar3 software with high reproducibility at an optimal annealing temperature of 54°C. The isolated DNA has successfully amplified with all seven primers designed from homological MYB gene sequences of rice genome. Polymerase chain reaction (PCR) has carried out in a final reaction volume of is 25 μ l, lowed concentrations of primer $(0.2\mu M)$ and Taq polymerase(1U). The amplified products were visualized as single compact bands of expected size of 400-500 base pairs. These sequences has successfully submitted to National Center for Biotechnology Information (NCBI) public database and obtained nucleotide Genbank Accession number (KF364950) and Locus (KF364950) for a public database. After the assigned accession number, there are 3 transcription factor MYB93-like mRNA sequences found from Prosopis alba which is a gum exudation tree species belong to its native ranges. This analysis used to study the genetic diversity of MYB genes in the arid trees. Moreover, it enables us to further understand the genetic diversity of MYB gene in relation to sequence analysis and find conserved molecular markers linked to secondarymetabolite regulatory mechanism. **Keyword:**Acacia senegal, mRNA, myeloblastosis and transcription factors

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I. Introduction

The potential of modern science to substantially benefit arid areas mainly deflation arid zone of India, which are at vulnerable due to the consequences of aeolian erosion and deliberated activities, leading to the biodiversity loss as well as extending the list of the threatened plants in the IUCN in a particular region. *Acacia senegal* one of the most dominant biomass producer of the sandy areas of the desert, belonging to (Fabaceae family and genus *Acacia*) locally known as Kummat, has remarkable adaptability to the adverse environmental conditions. Kummat, is a medium to large size with multiple utility like bio-fencing, food, fuel and nitrogen fixing tree distributed to arid and semi-arid regions of the country having fuel, metabolites of medicinal value and gums as its powerful implements in various applications. These plants has known to contain high levels and different types of secondary metabolites (gum Arabic), which are often responsible for their therapeutic and pharmacological activities. In modern technology, extraction of the plant definitelyboost up pharmaceutical production, urging the need for the plant conservation (Pirttilä*et. al*, 2001). Earlier studies also defined that *A. senegal*grown in various land forms and its girth class effect gum production (Pareek *et. al*, 2017 a & b).

The myeloblastosis (MYB) transcription factor gene family is found in all eukaryotes. In historical point of view, "Oncogene" v-MYB was the first MYB gene identified in avian myeloblastosis virus (Klempnauer*et. al*, 1982). Three v-MYB-related genes i.e., c- YB, A-MYB and B-MYB has subsequently identified in many vertebrates and implicated in the regulation of cell apoptosis, differentiation, and proliferation (Weston K, 1998). A homolog of mammalian c-MYB gene, i. e. Zea mays C1, who involved inregulation of anthocyanin biosynthesis, was the first MYB gene to be characterized in plants (Paz-Ares J *et. al*, 1987) Moreover, plants encode large number of MYB genes as compared to fungi and animals (Feller A *et.*

al, 2011; Yanhui C *et. al*, 2006). According to the number of adjacent MYB repeats, MYB transcription factors has further classified into four major groups, i.e. 1R-MYB, 2R-MYB, 3R-MYB and 4R-MYB. 1R-MYB, 2R-MYB, 3R-MYB and 4R-MYB contain one, two, three and four MYB repeats, respectively.

MYB transcription factors play a key role in plant development, secondary metabolism, hormone signal transduction, disease resistance and abiotic stress tolerance (Allan AC et. al, 2008; Cominelli E *et. al*, 2009). Several R2R3-MYB genes are involved in regulating responses to environmental stresses such as drought, salt, and cold (Agarwal M. *et. al*, 2006; Yanhui C *et. al*, 2006). Other important functions of MYBs have control of cell cycle regulation, secondary metabolism, meristem formation and thecellular morphogenesis (Ito M *et. al*, 2001; Haga N *et. al*, 2007; Ouyang *et. al*, 2007).

II. Material Methods

All the above find fact, homology-modeling search by using "MYB Acacia senegal" key word in NCBI public database. However, 1603 clusters of expressed transcripts found in UNI gene database. Out of which, 1320 expressed transcripts found in plants. We are extract the information from NCBI database and finally Rice genome (Riechmannet. al, 2000; Zeng et. al, 2007) were selected for homolog sequences of MYB transcription factor for A. senegal tree.

Primer designed from

leaves

Extract MYB gene sequences from Rice sequences from Rice sequences from Rice sequences from Rice Ten gene specific forward and reverse primer sequences has selected for PCR

Fig. 1 Step by step procedure to designed MYB gene specific primers through homology modeling approach for *Acacia Senegal*

- I. Sample fixation and pre-extraction washing: Leaves of medium size and age has used for isolation of DNA. Leaves has picked in the field, immediately frozen in liquid nitrogen (-196 °C) and ground to a fine powder using a pestle and mortar. Fixed samples were stored in the laboratory at -20 °C for up to an overnight.
- II. DNA isolation using Xcelgen Kit:Following standard the manufacturer's protocol, The Ultra violet (UV) absorbance has checked at 260 and 280 nm for determination of DNA concentration and purity. Purity of DNA has judged based on optical density ratio at 260:280 nm (Dellatropa, 1983; Saghai-Maroof*et. al.*, 1984). The DNA having ratio 0.8 to 2.0 has considered being of good purity. Quality of the extracted DNA has assessed in two ways. Firstly, the A260/A280 absorbance ration was determined using spectrophotometer (UV-Visible Elico spectrophotometer). The level of DNA purity was determined by the A260/A280 absorbance ratio.DNA purity has further determined by agarose gel electrophoresis. Agarose 0.8% (w/v) in 0.5X TAEi.e. a mixture of Tris base, acetic acid and Ethylenediaminetetraacetic acid (pH 8.0) buffer has used for submarine gel electrophoresis. Ethidium bromide (1%) has added @ 10µl /100ml. The wells has charged with 5µl of DNA preparations mixed with 1µl gel loading dye. Electrophoresis has carried out at 80V for 30 min at room temperature. DNA has visualized under UV using UV trans-illuminator. The DNA has used further for PCR (Fig.2).



Fig. 2 Genomic DNA of Acacia senegal form 1-7 and (M) ladder, resolved in 0.8% Agarose gel.

- III. Identification of MYB gene family in Rice: To identify "MYB transcription factor family genes", we searched and obtained genes annotated as MYB in public database (NCBI). Rice sequences obtain from NCBI server and selected corresponding MYB transcription factor gene location. Download all the relative sequences with referenced loci. Randomly selected query sequences for further primer design.
- IV. **Target Primer design:**Gene specific primers were designed using IDT PrimerQuest (http: //www. idtdna.com/scitools/applications/primerquest/default.aspx). We find total number of 10 forward and reverse primers, which are used as a homolog primer in *A. senegal* plant (Table 1).

Table 1. List of target gene specific primers of rice, which are used as a target primer of A.senegal MYB		
transcription factor sequence.		

Forward Primer Sequence	Reverse Primer Sequence
CCTCATGAACTACATTGCCTGCCA	AGTTCTTGATCTCGTTGTCCGTCCT
CGCAAGCTCAAGCTCAAACAGCAA	TTGGTGCTGGTCCTCATGATGTTG
TTGGCAACAAGTGGGCAGCAATAG	AGGGTTGTAGTAGTTTGGGCTGGT
ACACCAAGCAAACTCCATGCACAG	AGTTCATCGCTTCTTTGCTGCCAC
ATCAGGGCTCAATAGGACAGGCAA	TCCTGGGCTTTCTTCCTCATGTGT
CAAAGCAACTCCCACCATCAGCAA	ATAGTTCCTTCGGTGGCACCTTCA
ATTGGAGAGGTGTGGCAAGAGTTG	GGAGCTTCTTCTTGAGGCACGAGT
AACGACGTCAAGAACCACTGGAAC	TGCAGGAAGTCGTCGCAGGTGTA

V. Polymer Chain Reaction: PCR was carried out in a final reaction volume of 25 µl in 200 µl capacity thinned wall PCR tube in Eppendorf Thermal Cycler. PCR tubes containing the mixture has tapped gently and spin briefly at 10,000 rpm. The PCR tubes with all the components has transferred to thermal cycler. The primer (forward and reverse) used for an amplification of genomic DNA (Table 2&3).

Table 2 Composition of reaction mixture for T CK		
Components	Quantity	Final
DNase-RNase free water	7.5µl	
2X PCR master mix (MBI Fermenas	12.50 µl	1x
Forward Primer (10P/ µl	1.00 µl	10 Pmole
Reverse Primer (10 Pmole/ µl	1.00 µl	10 Pmole
Diluted DNA (30 ng/ µl)	3 µl	
Grand Total	25.0 µl	

 Table 2 Composition of reaction mixture for PCR

Table 3 Steps and condition	ons of thermal cycling for PCR
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Steps	Temperature	Time	Cycles
Initial Denaturation	95°C	2 min	1
Final Denaturation	94°C	30 Sec	30
Annealing	50-54°C	30 Sec	

Extension	72°C	90 Sec	
Final Extension	72°C	10 min	1

VI. Visualization the PCR product: To confirm the targeted PCR amplification, 5 μl of PCR product from each tube was mixed with 1 μl of 6X gel loading dye and electrophoreses on 1.2 % agarose gel containing ethidium bromide (1 per cent solution @10 μl/100 ml) at constant 5V/cm for 30 min in 0.5 X TAE buffer. The amplified product has visualized as a single compact band of expected size under UV light and documented by gel documentation system (Fig. 3).



Fig 3. L1=1 KB Ladder (starting from bottom 500BP, 1KB, 2KB, 3KB, 4KB, 5KB, 6KB, 7KB, 8KB) and ladder L2=100 BP (starting from bottom-100BP, 200BP, 300BP, 400BP, 500BP, 600BP, 700BP, 800BP, 900BP, 1KB) showing amplification of MYB gene from *Acacia senegal*.

VII. Sequencing of Purified DNA

Sequencing of Gene Segment: The concentration of the purified DNA has determined and has subjected to automated DNA sequencing on ABI 3730xl Genetic Analyzer (Applied Biosystems, USA). Sequencing has carried out using BigDye® Terminator v3.1 Cycle sequencing kit following manufacturer's instructions.

Cycle Sequencing:Cycle sequencing has performed following the instructions supplied along with BigDye® Terminator v3.1 Cycle Sequencing Kit. The reaction has carried out in a final reaction volume of 20μ l using 200μ l capacity thinned wall PCR tube. The cycling protocol (Table 4) has designed for 25 cycles with the thermal ramp rate of 1°C per second.

Step*	Temperature	Time
Denaturation	96°C	10 sec
Annealing	53°C	5 sec
Extension	60°C	4 min

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Table 4 Cycling	g protocol for see	quencing reaction

* Repeat step 1 to 3 for 25 cycles

The contents hasmixed on a shaker for 30 minutes at 300xg. Eluted PCR products has placed in a sample plate and covered with the septa. Sample plate has heated at 95°C for 5 min, snap chilled and loaded into autosampler of the instrument.

Electrophoresis and Data Analysis:Electrophoresis and data analysis has carried out on the ABI 3730xl Genetic Analyzer using appropriated Module, Basecaller, yeset/Primer and Matrix files.

Submission of MYB Transcroptom factor sequence in NCBI public database:Submission of MYB transcroptom factor sequence in NCBI public database: To find out ORF in candidate sequences obtained from sequencer, sequences has input to ORF finder. Systematically methods apply to find out best sequences. The most important source of new data for GenBank® is direct submissions. The submission wizard helps to upload the sequences in GenBank. After successfully submitted sequences, primary accession no obtained from GenBank. Finally, one sequences has identified and allot accession number i.e. KF364950 (Pareek *et. al*, 2013).

VIII. Result and discussion

A major problems in isolation of high quality DNA from mature tree leaves especially in arid region trees, however it still an important issuesbecause it will contain high amount of phenolic compounds (Katterman and Shattuck, 1983) and as well as polysaccharides (Barzegariet. al., 2010; Li et. al., 2010; Varma et. al., 2007). During experiments, we noticed that leaves size and age, and locality are very important traits of isolation of PCR usable DNA (Kim CS et. al., 1999; Khanujaet. al., 1999; Sabloket. al., 2009). Medium size and age of leaves are suitable for isolation high quality DNA. It is remarkably that no any sequences in NCBI data base related to A. senegal MYB gene. It very difficult to compare homologue sequences from model plant (Rice), with the help of various tools. Conserved domains of MYB gene related sequence selected for designing primers. In this study, we find out seven MYB gene specific novel sequences. Out of which, only one sequence were successfully submittedto GenBank and published with GeneBank identifier ID (KF364950); Locus (KF364950) and rest six sequences were submitting to ESTs database because these sequences were very short in length (range less than 200). The identify nucleotide sequences similar to the KF364950, a blast results revealed that 3 sequences of Prosopis alba tree species were identical in relation to its taxonomical approaches. Out of 101 identical organisms, in which 3 sequences of Prosopis alba transcription factor MYB93-like mRNA which identical locus are LOC114719063,LOC114716567 and LOC114729076 found with query cover range 48% to 54% identical. This study indicated that more precisely study will need for arid tree species. It is interestingly that only one sequence was found till date related to A. senegalMYB gene partial sequence in NCBI public database.

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Competing interests

The authors declare that they have no competing interests.

References

- [1]. Agarwal M, Hao Y, Kapoor A, Dong CH, Fujii H, Zheng X and Zhu JK. (2006). A R2R3 type MYB transcription factor is involved in the cold regulation of CBF genes and in acquired freezing tolerance. *J BiolChem*, 281:37636–37645.
- [2]. Allan AC, Hellens RP and Laing WA.(2008). MYB transcription factors that colour our fruit. Cell, 13:99–102.
- [3]. Barzegari, A., Vahed, SZ., Atashpaz, S., Khani, S. andOmidi, Y.(2010). Rapid and simple methodology for isolation of high quality genomic DNA from coniferous tissues (Taxusbaccata). *Mol. Biol. Rep.* 37, 833–837.
- [4]. Cominelli E and Tonelli C. (2009). A new role for plant R2R3-MYB transcription factors in cell cycle regulation. *Cell Res*, 19:1231–1232.
- [5]. Feller A, Machemer K, Braun EL andGrotewold E.(2011). Evolutionary and comparative analysis of MYB and bHLH plant transcription factors. *Plant J*, 66(1):94–116.
- [6]. Haga N, Kato K, Murase M, Araki S, Kubo M, Demura T, Suzuki K, Muller I, Voss U, Jurgens G and Ito M. (2007). R1R2R3-MYB proteins positively regulate cytokinesis through activation of KNOLLE transcription in Arabidopsis thaliana. *Development*, 134:1101–1110.
- [7]. Ito M, Araki S, Matsunaga S, Itoh T, Nishihama R, Machida Y, Doonan JH and Watanabe A.(2001). G2/M-phase-specific transcription during the plant cell cycle is mediated by c-MYB-like transcription factors. *Plant Cell*, 13:1891–1905.
- [8]. Katterman FRH and Shattuck VI. (1983). An effective method of isolation from the mature leaves of Gossypium species that contain large amounts of phenolics, terpeniodes and tannins. *Preparative Biochemistry* 13: 347-359.
- [9]. Khanuja SPS, Shasany AK, DarokarMP andKumar S. (1999) Rapid isolation of DNA from dry and freshsamples of plants producing large amounts of secondary metabolites and essential oil. *Plant Molecular Biology Reporter* 17: 1-7.
- [10]. Kim CS, Lee CH, Shin JS, Chung YS and Hyung NI. (1997). A simple and rapid method of isolation of high quality genomic DNA from fruit trees and conifers using PVP. Nucleic Acids Research 25: 1085–1086.
- [11]. Klempnauer KH, Gonda TJ and Bishop JM. (1982). Nucleotide sequence of the retroviral leukemia gene v-myb and its cellular progenitor c-MYB: the architecture of a transduced oncogene. *Cell*, 31:453–463.
- [12]. Li, J.F., Li, L. and Sheen, J.(2010). Protocol: a rapid and economical procedure for purification of plasmid or plant DNA with diverse applications in plant biology. Plant Methods 6, 1. doi:10.1186/1746-4811-6-1. Dellaporta SL, Wood J, Hicks JB. 1983. A plant DNA minipreparation: version II. *Plant Molecular Biology Reporter* 1: 19-21.
 [13]. Ouyang S, Zhu W, Hamilton J, Lin H, Campbell M, Childs K, Thibaud-Nissen F, Malek RL, Lee Y, Zheng L, Orvis J, Haas B,
- [13]. Ouyang S, Zhu W, Hamilton J, Lin H, Campbell M, Childs K, Thibaud-Nissen F, Malek RL, Lee Y, Zheng L, Orvis J, Haas B, Wortman J and Buell CR.(2007). The TIGR rice Ggenome annotation resource: improvements and new features. *Nucleic Acids Res*, 35:D883–D887.
- [14]. PareekK, Kalia RK, Tewari JC and Ram M. (2013). Acacia senegal MYB transcription factor mRNA, partial cds (Accession number: KF364950; GI 553041288): NCBI Submission (https://www.ncbi.nlm.nih.gov/nuccore/553041288/)
- [15]. PareekK, TewariJC, Shiran K, GaurMK, SharmaA and Chaudhary, V (2017 a). Effect of different girth classes on gum Arabic production from Acacia senegal in arid western Rajasthan. IOSR Journal of Environmental Science, Toxicology andFood Technology (IOSR-JESTFT), 11, 9, 12–16.
- [16]. PareekK, Tewari, JC, Shiran K, Gaur MK, Bishnoi PR, Sharma A and Chaudhary V (2017 b). Gum production on different land forms in arid western Rajasthan. *IOSR Journal of Agriculture and Veterinary Science* (IOSR-JAVS), 10, 9, 14–17.
- [17]. Paz-Ares J, Ghosal D, Wienand U, Peterson P and Saedler H. (1987). The regulatory c1 locus of Zea mays encodes a protein with homology to MYB oncogene products and with structural similarities to transcriptional activators. *EMBO J*, 6:3553–3558.
- [18]. Pirttilä AM, Hirsikorpi M, Kamarainen T, Jaakola L andHohtola A. (2001). DNA isolation methods for aromatic and medicinal plants. *Plant Molecular Biology Reporter* 19: 273a–273e.

- [19]. Riechmann JL, Heard J, Martin G, Reuber L, Jiang C, Keddie J, Adam L, Pineda O, Ratcliffe OJ, Samaha RR, Creelman R, Pilgrim M, Broun P, Zhang JZ, Ghandehari D, Sherman BK and Yu G.(2000). Arabidopsis transcription factors: genome-wide comparative analysis among eukaryotes. Science 2000, 290:2105–2110.
- [20]. Sablok G, Gahlot P, Gupta AK., Pareek K and ShekhawatNS. (2009). Extraction of PCR- usable DNA from trees adapted to arid environment. *Plant Omics Journal 2(3), 103-109.*
- [21]. Saghai MM. A, Soliman KM, JoregensenRA and Allard RW. (1984). Ribosomal DNA spacer-length polymorphisms in barley: Mendelianinheritence, chromosomal location, and population dynamics. *Proceedings of the National Academy of Sciences* 81:8014-8018.
- [22]. Varma A, Padh H and Shrivastava N.(2007). Plantgenomic DNA isolation: an art or a science. Biotechnology Journal 2: 386–392.
- [23]. Weston K.(1998).Myb proteins in life, death and differentiation. *CurrOpin Genet Dev*, 8:76–81.
 [24]. Yanhui C, Xiaoyuan Y, Kun H, Meihua L, Jigang L, Zhaofeng G, Zhiqiang L, Yunfei Z, Xiaoxiao W, Xiaoming Q, Yunping S, Li
- Z, Xiaohui D, Jingchu L, Xing-Wang D, Zhangliang C, Hongya G and Li-Jia Q. (2006). The MYB transcriptionfactor superfamily of Arabidopsis: expression analysis and phylogenetic comparison with the rice MYB family. *Plant MolBiol*, 60:107–124.
- [25]. Zeng H, Luo L, Zhang W, Zhou J, Li Z, Liu H, Zhu T, Feng X and Zhong Y.(2007) .PlantQTL-GE: a database system for identifying candidate genes in rice and Arabidopsis by gene expression and QTL information. *Nucleic Acids Res*, 35:D879–D882.

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